

University at Albany, State University of New York

Scholars Archive

Chemistry

Honors College

12-2021

Raman Spectroscopy for Forensic Identification of Body Fluid Traces: Method Validation for Potential False Negatives Caused by Blood-Affecting Diseases

Niara Nichols

The University at Albany community has made this article openly available.

Please share how this access benefits you.

Follow this and additional works at: https://scholarsarchive.library.albany.edu/honorscollege_chem



Part of the [Chemistry Commons](#)

**Raman Spectroscopy for Forensic Identification of Body Fluid Traces:
Method Validation for Potential False Negatives Caused by Blood-Affecting Diseases**

An honors thesis presented to the
Department of Chemistry,
University at Albany, State University of New York
in partial fulfillment of the requirements
for graduation with Honors in Chemistry
and
graduation from The Honors College

Niara A. Nichols

Research Mentor: Igor K. Lednev, Ph.D.

December 2021

Abstract

Two critical issues in forensic science are identifying body fluid traces found at crime scenes and preserving them for DNA analysis. However, the majority of current biochemical tests for body fluid identification, which are applicable at the crime scene, are presumptive and destructive to the sample. Raman Spectroscopy provides a suitable alternative to current methods as a nondestructive, confirmatory, and potentially in field method. Our laboratory has developed a chemometric model for the identification of five main body fluids using Raman spectroscopy. This model was developed using samples obtained from healthy donors. Thus, it is of most importance for the forensic application of the method to validate its performance for donors with diseases that might affect the biochemical composition of body fluids. In this study, the developed method was validated using peripheral blood samples acquired from donors with Celiac Disease, Sickle Cell Anemia, and Type 2 Diabetes. It was shown that the method correctly identified all samples as peripheral blood indicating that no false positives could occur because the blood traces were originated from donors suffering from the diseases.

Keywords: *Raman spectroscopy, Chemometric, Bloodstain, False negatives, Body fluids*

Acknowledgements

I would like to thank those who have made the completion of this thesis possible. I would like to thank Casey Kohler who helped me get involved with research at the start of my undergraduate career and has continued to encourage me these past three and a half years. I would like to thank the members of my lab for their continued support, notably Marisia Fikiet for mentoring me at the beginning of this project. I would especially like to thank Dr. Igor Lednev, my research advisor, for accepting me into his lab, mentoring me for these past three and a half years, and helping me write and revise this thesis.

I would also like to thank the National Institute of Justice, Office of Justice Programs, U.S. Department of Justice for funding this project with Award No. 2017-DN-BX-0135.

Finally, I would like to thank my parents who have done nothing but encourage me in everything I pursue. None of the work I have done thus far would be possible without their love and support.

List of Figures

Figure 1	6
Figure 2	7

Table of Contents

Abstract	ii
Acknowledgment	iii
List of Figures	iv
Introduction	1
Methods	4
A. Samples.....	4
B. Instrumentation	4
C. Statistical Analysis.....	5
Results and Discussion	5
Conclusions	9
References	11

Introduction

The identification of body fluid stains found at crime scenes is a major component of forensic investigations. Knowledge of the identity and origin of such evidence can provide important context about a crime. Further, body fluids can provide important information about potential victims and suspects, such as race or gender [1, 2]. Most notably, DNA evidence can be collected from body fluid stains and used for personal identification [3]. Recent work has also looked into using body fluid stains to deduce the time since their deposition, and thus, the approximate time of the crime [4]. Considering the wealth of information that can be extracted from a single body fluid stain, it is important that suitable body fluid identification tests are accessible to analyst. Ideally, such a test could be universally applied to all body fluids, used in the field, and wouldn't destroy the often, little amount of sample found at the crime scene. Further, such a test should be confirmatory, meaning it doesn't yield false negatives (a negative result that is actually positive) or false positives (a positive result that is actually negative).

Methods that forensic analyst currently use contain many of these characteristics and offer a robust analysis of body fluids, however, they often fall short of the ideal. Presumptive chemical tests that can be used in the field, such as Kastle Meyers test, are quite sensitive. However, they aren't awfully specific and can yield false positive results for a variety of common household substances and forensically relevant materials like denim, as well as false negatives [5, 6]. Thus, confirmatory analysis is required in the lab to ensure that blood is present. Confirmatory tests for blood like the Takayama and Teichman test are effective in their selective identification of blood but are destructive in nature [7]. More often, forensic analyst may go directly for DNA analysis to confirm the presence of human blood. While this method is direct

and confirmatory, it must be conducted in a lab setting as opposed to in the field and can be timely [3].

Raman spectroscopy can potentially serve as an alternative to these methods and remedy these shortcomings. It has a number of uses in forensic science [8]. Pairing Raman spectroscopy with chemometric analysis can allow for the confirmatory identification of body fluids, even in the presence of some contaminants [9, 10]. The advantages don't stop here. Raman spectroscopy is non-destructive; thus, it won't consume the trace amount of sample being analyzed. Further, this method can be universally applied to several body fluids, as opposed to using a specific test to search for each type of body fluids [9, 11]. This universal approach can save both money and time. Finally, recent advancements have allowed for the creation of portable Raman spectrometers, meaning that there is a possibility for use in the field.

Having acknowledged the many advantages provided by the use of Raman spectroscopy for the analysis of body fluid stains, our laboratory has developed a novel method that can identify the five main body fluids (peripheral blood, semen, saliva, sweat, and vaginal fluid) from their Raman spectra using a machine learning model [9]. This method, outlined in Muro et al., 2016, uses a support vector machine discriminant analysis model with genetic algorithm. This method was shown to have 100% success in the analysis of external validation samples.

While this method has already been shown to have remarkable promise for the more efficient analysis of body fluid stains, it still needs thorough validation before practical application by law enforcement agencies. Notably, the model used in this method was constructed using body fluid spectra from samples that came from healthy donors. There are a host of diseases that can affect the biochemical composition of body fluids, which in turn could

affect their Raman spectra, and the ability of the model to identify the body fluid correctly. Thus, to ensure the forensic relevance of this method, potential false negatives arising from diseases that could affect the biochemical composition of a body fluid must be evaluated.

In this paper, the effect of blood-affecting diseases on the ability of the model to identify peripheral blood was evaluated. The chosen blood-affecting diseases have already been shown to pose a potential risk for false negatives due to their Raman spectra's dependence on the disease. While this is useful for disease diagnostics, it can complicate forensic analysis. The first blood-affecting disease, Celiac disease, is an autoimmune disorder where in individual's small intestine will damage themselves if they don't remove gluten from their diet. This disorder affects 1 in 100 people world-wide [12]. Ralbovsky & Lednev have shown that blood from donors with Celiac Disease differs from that from healthy donors on a gluten-free diet [12].

The Raman spectra of blood has also been shown to be affected by whether or not an individual has Sickle Cell Anemia. This genetic disorder is characterized by abnormal hemoglobin, such as Hemoglobin S, which arises due to the replacement of glutamic acid with valine in the hemoglobin β amino acid chain [13]. As a result of the abnormal hemoglobin, the red blood cell forms a sickle shape. This can cause a variety of complications such as high blood pressure or even heart failure [14]. The changes seen in the Raman spectra of Sickle Cell Anemia blood can be explained by differences in hemoglobin. As it dominates the Raman spectra of blood, differences in hemoglobin can have such an effect on the overall Raman spectra [13, 14].

Type 2 Diabetes is a condition characterized by abnormally high glucose levels in the blood due to scarce or ineffective insulin, which can lead to severe complications such as organ failure [15]. This particular type of diabetes is the most prevalent form with 8.6% of American

adults having Type 2 diabetes compared to 0.55% of the same population having Type 1 diabetes [16]. The Raman spectra of blood from donors with Diabetes has also been shown to differ from healthy controls. This could be due to several biomarkers of the disease, such as Hemoglobin A1c [15].

In this study, the validation of the Raman spectroscopic method for the identification of body fluid traces was validated with respect to potential false negatives caused by blood effecting diseases; Celiac Disease (CD), Sickle Cell Anemia (SCA), and Type 2 Diabetes (D2).

Methods

A. Samples

Whole blood samples from 10 donors with CD, 3 donors with SCA, 4 donors with D2, and 1 donor with both SCA and D2 were purchased from BioIVT, Inc. (Westbury NY). The donors were chosen such that there was variety in age, sex, and race. This is with the exception of the race of SCA donors, who were all African American. The samples were stored in a -80°C freezer until use. Upon use, the samples were thawed, and 5 μ L of the whole blood was deposited onto an aluminum covered glass slide. The slide was stored in a petri dish to prevent contamination and allowed to dry overnight.

B. Instrumentation

The Raman spectra of the blood samples were collected using an inVia Raman spectrometer (Renishaw, Inc. Hoffman Estate, IL) with WiRE 3.2 software and a Lecia research grade microscope. Prior to sample collection, the spectrum of a silicon standard was collected to ensure the instrument was properly calibrated. A 20x objective was used to focus the laser beam

onto the samples. A 785 nm excitation wavelength was used at a laser power of 5%. For each spectrum, 20 ten-second accumulations were collected over a 300 – 1800 cm^{-1} range. 10-20 spectra were collected from different points within the same bloodstain using automatic mapping to account for intrasample variability. These parameters aligned with those used to record the Raman spectra of healthy blood used to construct the model developed in Muro et al., 2016.

C. Statistical Analysis

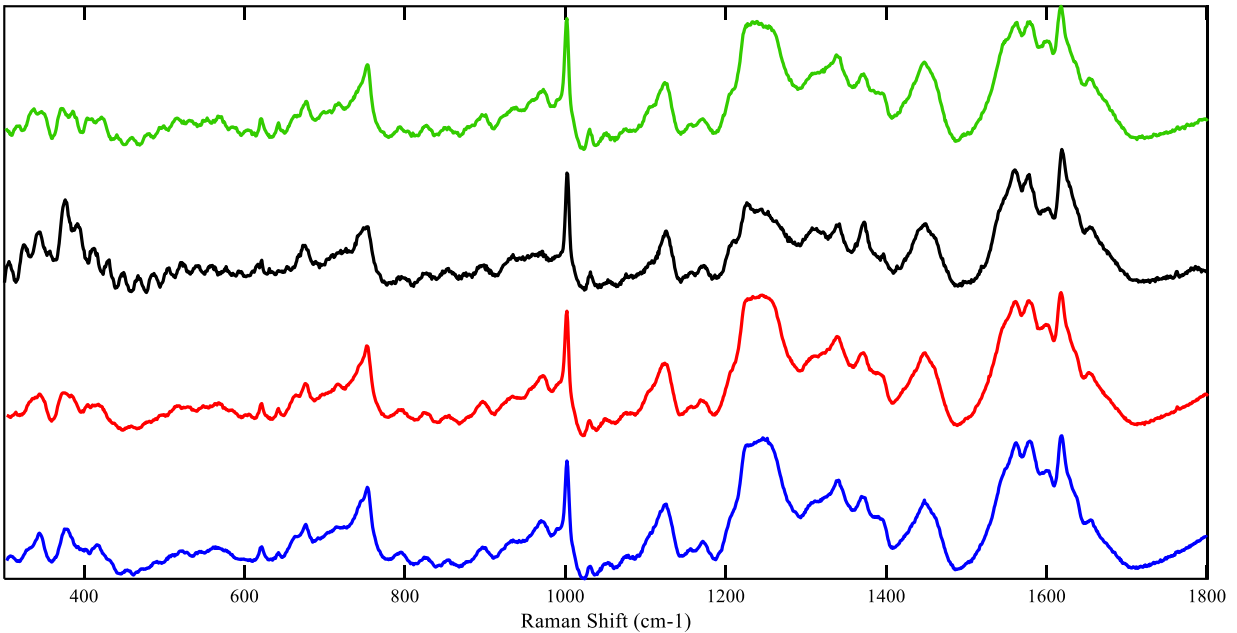
Cosmic rays were first removed from the Raman spectra of the blood samples in the WIRE 3.2 software. After, the spectra were preprocessed in MATLAB (MathWorks inc., Natick, MA) using the PLS Toolbox extension (Eigenvector Research, Wenatchee, WA). In accordance with the preprocessing used in Muro et al., 2016, the spectra were first baseline corrected using automatic weighted least squares, 5th order polynomial. Then the spectra were normalized by total area. After preprocessing, each group of spectra were analyzed using the Support Vector Machine Discriminant Analysis (SVMDA) model with Genetic Algorithm (GA) created in Muro et al., 2016. The model predicted which body fluid class (if any) the spectra belonged to and displayed it in a confusion matrix along with the actual body fluid class of the spectra.

Results and Discussion

This study sought to further validate a previously developed body fluid identification model based on Raman spectroscopy by investigating the potential for false negatives caused by blood-affecting diseases. In this study, 213 Raman spectra were collected from 18 peripheral bloodstains obtained from donors with various blood-affecting diseases.

Figure 1

Average Spectra by Disease Group

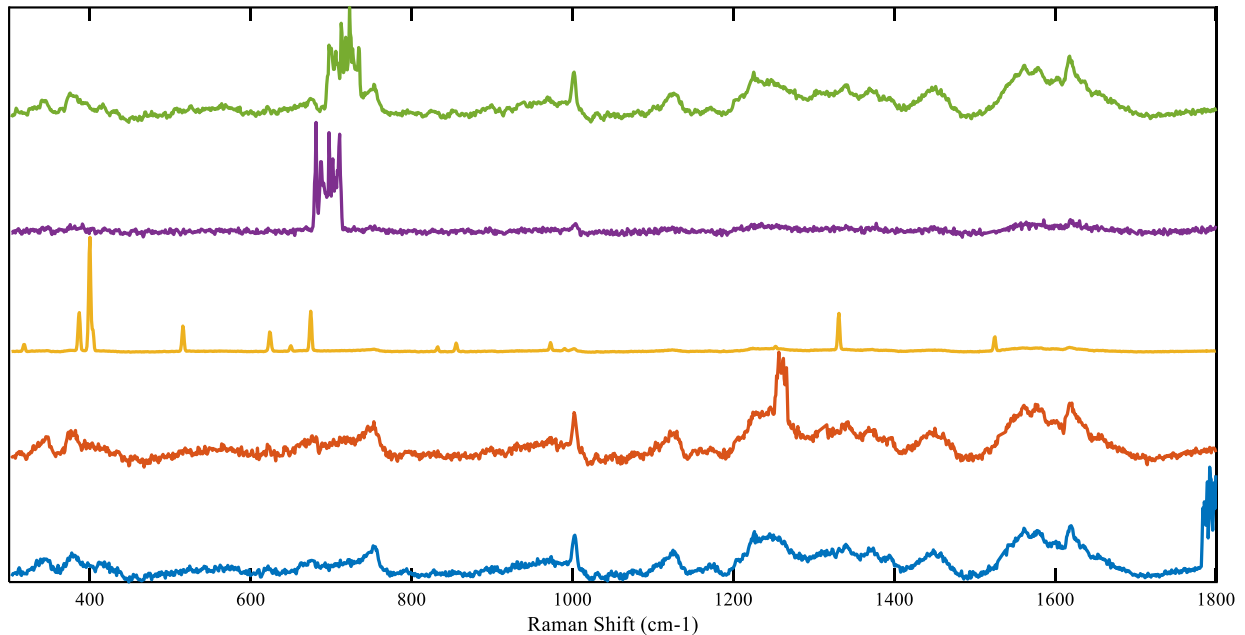


Note. This figure shows the average preprocessed Raman spectra of peripheral bloodstains for the healthy peripheral blood used to test the study's protocol (black) and each of the disease groups (Celiac Disease – blue, Sickle Cell Anemia – green, Type 2 Diabetes – red).

Peripheral blood from donors with CD, SCA, and D2 were evaluated in this study. The average preprocessed Raman spectra for each disease are displayed in Figure 1, alongside the average preprocessed Raman spectra collected from healthy controls that was used to test this study's protocols. The spectra exhibit remarkable similarity upon visual examination. Still, given the subjective and non-computational nature of visually evaluating spectra, analysis using the statistical model is necessary to ensure that the blood-affecting diseases won't contribute to false negatives.

Figure 2

Excluded Celiac Disease Raman Spectra



Note. This figure shows the Celiac disease Raman spectra excluded from the analysis based on visual inspection.

Prior to analysis using the SVMDA model, the Raman spectra were preprocessed in the same manner that the spectra used to build the model were. Then, each disease group was evaluated by the model. While 100 CD spectra were collected, 95 were preprocessed and analyzed using the model, as 5 of the Raman spectra were excluded from the data set. Figure 2 shows the five spectra that were excluded based on visual inspection. These spectra exhibited excessive noise, or exorbitant amounts of cosmic rays. The obstruction by the noise/cosmic rays was evident from the visual examination and justified the removal of these spectra from further consideration, leaving 95 spectra to be analyzed by the model. Of the 95 CD spectra analyzed, all 95 were correctly identified as peripheral blood.

For the other two diseases evaluated, SCA and D2, no spectra were excluded from preprocessing and analysis with the model. Thus, 52 SCA spectra and 75 D2 spectra were analyzed. It should be noted that one sample, for which 14 spectra were collected, is double counted for these two disease groups, as the donor had both diseases. All of the SCA and D2 spectra were correctly identified by the model as peripheral blood.

The model was successful in identifying the 208 spectra resulting in 100% accuracy. The correct classification of all the spectra means that the model exhibited no false negatives despite analyzing samples from donors with blood-affecting diseases. These findings suggest that the analyzed diseases don't pose a risk for causing false negatives. This could be attributed to the fact that at the excitation wavelength used for analysis, 785 nm, the Raman spectra of blood is dominated by hemoglobin. Some of the diseases do influence hemoglobin, such as SCA which is caused by the atypical hemoglobin S molecule. However, the results suggest that the composition of these hemoglobin molecules and healthy hemoglobin are largely the same. While the difference between the two may be enough to result in functional differences and differentiation when a model is trained to do so, their Raman spectra are similar enough to still allow for the analyzed blood-affecting disease blood to be correctly identified.

Conclusion

Body fluid traces remain a crucial piece of evidence for forensic investigations, making the identification and preservation of this type of evidence of the utmost importance. While current methods for the identification of body fluids often require further testing and destroy the evidence in question, Muro et al. has taken a step towards the development of a universal, non-destructive, and confirmatory body fluid identification method. This method, which uses Raman spectroscopy paired with SVM and GA, has proven to be highly effective for the identification of the five main body fluids (peripheral blood, semen, vaginal fluid, sweat, and saliva). As samples from healthy controls were used to construct and validate the method, to ensure its confirmatory nature, the potential for false negatives caused by blood-affecting diseases was evaluated. It was investigated whether the method could correctly identify peripheral blood that came from donors who have CD, SCA, and D2. The method correctly classified all the analyzed peripheral blood spectra, thus achieving 100% accuracy. This finding suggests that the presence of the blood-affecting diseases don't result in false negatives for this method.

The findings of this study further demonstrate Raman spectroscopy's potential to be used as a non-destructive and confirmatory method for the forensic identification of body fluids. In order for the method that was validated in this study to be considered confirmatory, it would need to have little to no risk of false negatives. The findings presented in this study support the confirmatory nature of Raman spectroscopy paired with chemometric analysis as the presence of blood-affecting diseases didn't result in false negatives. Still, before this method can be widely used by analyst in the field, there are other matters worth investigating. Other body fluid affecting diseases are worth examining, especially for the other main body fluids. Thus far, a

2018 study by Fikiet et al. examined the potential for azoospermia to cause false negatives for semen identification [17]. Other potential sources of false negatives should also be investigated, such as the reagents used for the preliminary detection of bloodstains at the scene of the crime, like luminol or fluorescein. Such investigations would push this method one step closer to becoming the non-destructive and confirmatory universal body fluid identification method forensic science needs.

References

1. Muro, C.K., L. de Souza Fernandes, and I.K. Lednev (2016) Sex determination based on Raman spectroscopy of saliva traces for forensic purposes. *Analytical Chemistry*, **88**(24), 12489-12493.
2. Mistek, E., et al. (2016) Race differentiation by Raman spectroscopy of a bloodstain for forensic purposes. *Analytical Chemistry*, **88**(15), 7453-7456.
3. McDonald, J. and D.C. Lehman (2012) Forensic DNA analysis. *Clinical Laboratory Science*, **25**(2), 109.
4. Doty, K.C., G. McLaughlin, and I.K. Lednev (2016) A Raman “spectroscopic clock” for bloodstain age determination: the first week after deposition. *Analytical Bioanalytical Chemistry*, **408**(15), 3993-4001.
5. Lalonde, W. and J.S. Millman (2019) Case study: Loss of Kastle-Meyer test specificity on jeans. *Science & Justice*, **59**(3), 359-361.
6. Tobe, S.S., N. Watson, and N.N. Daeid (2007) Evaluation of six presumptive tests for blood, their specificity, sensitivity, and effect on high molecular-weight DNA. *Journal of Forensic Sciences*, **52**(1), 102-109.
7. Virkler, K. and I.K. Lednev (2009) Analysis of body fluids for forensic purposes: from laboratory testing to non-destructive rapid confirmatory identification at a crime scene. *Forensic Science International*, **188**(1-3), 1-17.
8. Khandasammy, S.R., et al. (2018) Bloodstains, paintings, and drugs: Raman spectroscopy applications in forensic science. *Forensic Chemistry*, **8**, 111-133.
9. Muro, C.K., et al. (2016) Forensic body fluid identification and differentiation by Raman spectroscopy. *Forensic Chemistry*, **1**, 31-38.

10. Sikirzhyskaya, A., et al. (2013) Forensic identification of blood in the presence of contaminations using Raman microspectroscopy coupled with advanced statistics: effect of sand, dust, and soil. *Journal of Forensic Sciences*, **58**(5), 1141-1148.
11. Vyas, B., L. Halamkova, and I.K. Lednev (2020) A universal test for the forensic identification of all main body fluids including urine. *Forensic Chemistry*, **20**, 100247.
12. Ralbovsky, N.M. and I.K. Lednev (2021) Analysis of individual red blood cells for Celiac disease diagnosis. *Talanta*, **221**, 121642.
13. Bueno Filho, A.C., et al. (2015) Raman spectroscopy for a rapid diagnosis of sickle cell disease in human blood samples: a preliminary study. *Lasers in Medical Science*, **30**(1), 247-253.
14. da Silva, W.R., L. Silveira, and A.B. Fernandes (2020) Diagnosing sickle cell disease and iron deficiency anemia in human blood by Raman spectroscopy. *Lasers in Medical Science*, **35**(5), 1065-1074.
15. Ralbovsky, N.M. and I.K. Lednev (2021) EXPRESS: Vibrational Spectroscopy for Detection of Diabetes: A Review. *Applied Spectroscopy*, 00037028211019130.
16. Bullard, K.M., et al. (2018) Prevalence of diagnosed diabetes in adults by diabetes type—United States, 2016. *Morbidity Mortality Weekly Report*, **67**(12), 359.
17. Fikiet, M.A. and I.K. Lednev (2019) Raman spectroscopic method for semen identification: Azoospermia. *Talanta*, **194**, 385-389.